Eco-evolutionary dynamics of cooperative antimicrobial resistance in time-varying environments with spatial structure

<u>Lluís Hernández-Navarro</u>, Matthew Asker, Kenneth Distefano, Alastair M. Rucklidge, Uwe C. Täuber, and Mauro Mobilia

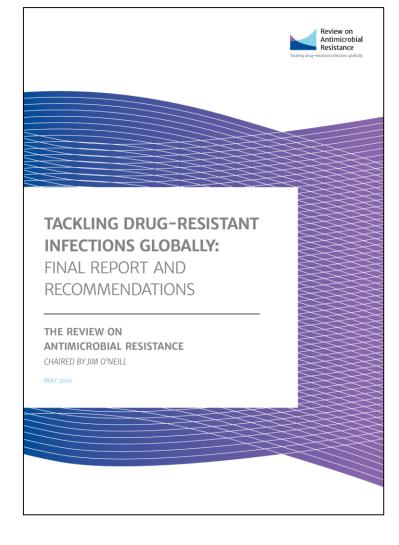
ANR meeting 28-29/11/24

КК

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Some context on AntiMicrobial Resistance (AMR)



• 10⁶ deaths/year



• 20 billion \$ US health excess costs

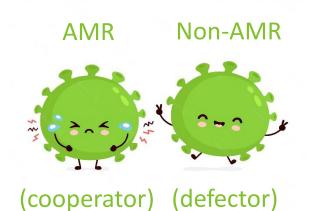
 10⁷ deaths/year by 2050 (more than cancer)



HEALTH

J. O'Neill, *Tackling drug-resistant infections globally: final report and recommendations*, Review on Antimicrobial Resistance (2016)

AMR: cooperative (Public Good) and comes with an extra metabolic cost



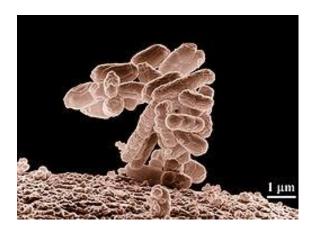
Microorganisms live in ecologically dynamic environments.



Main question(s)

In dynamic environments, when do resistant and sensitive strains coexist?

Otherwise, which strain dominates?



Model

(inspired by experiments with E. Coli, ampicillin, and pSEVA121 at ICL)

E. Coli

Imperial College London

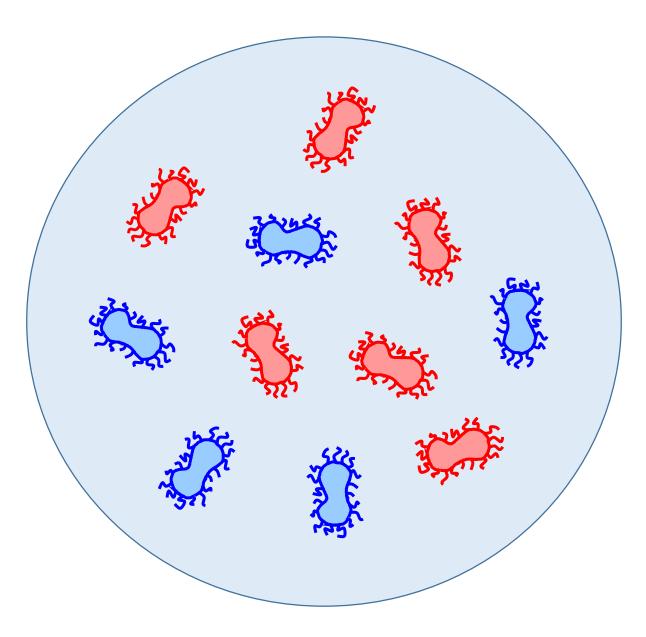


PhD candidate Said Muñoz Montero



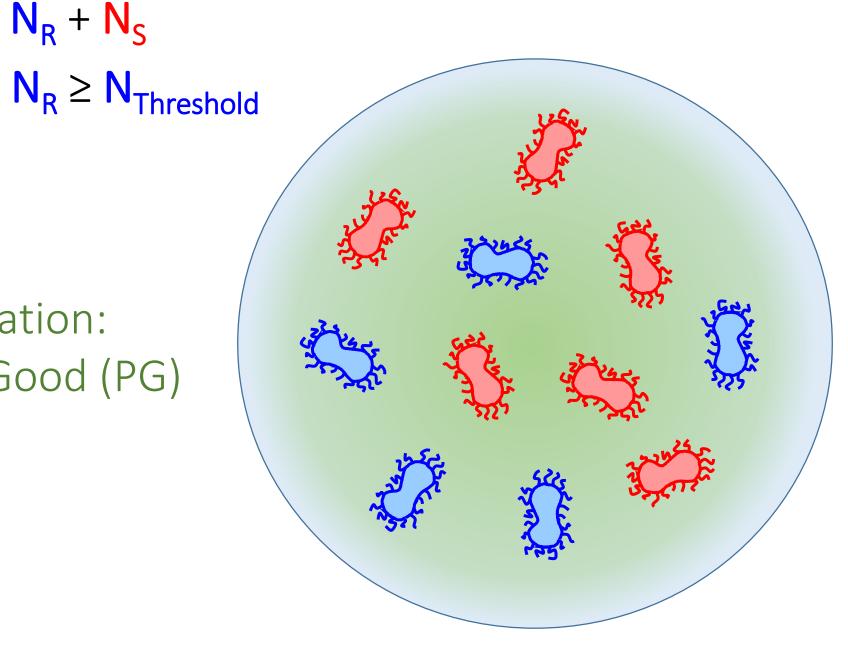
Prof. José Jiménez

 $N = N_{R} + N_{S}$



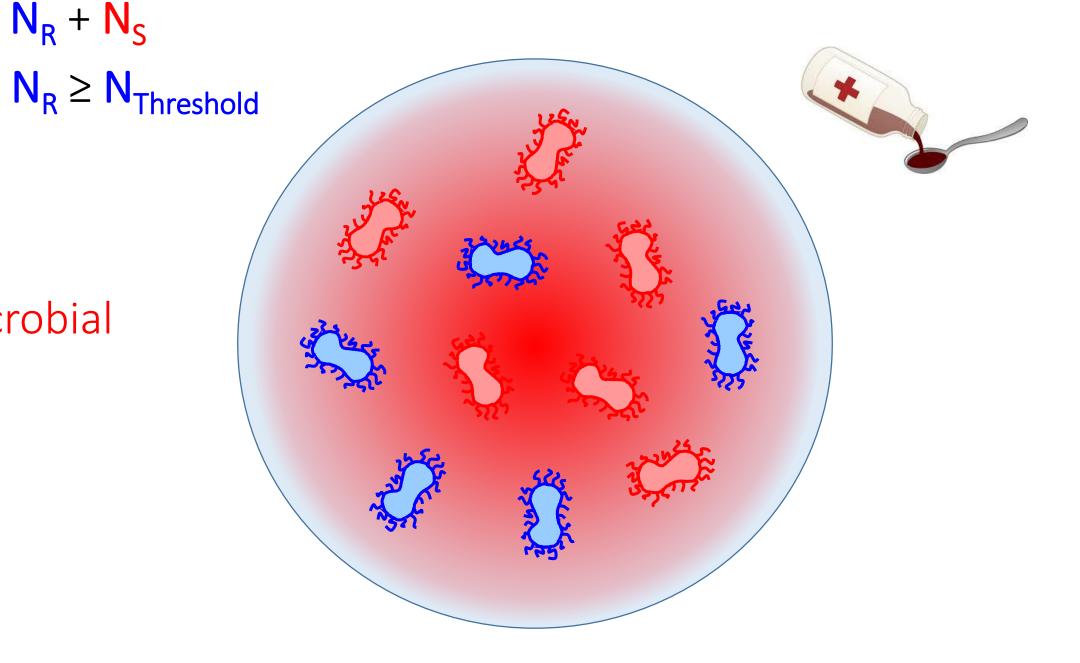
Cooperation: Public Good (PG)

 $N = N_{R} + N_{S}$



Antimicrobial drug

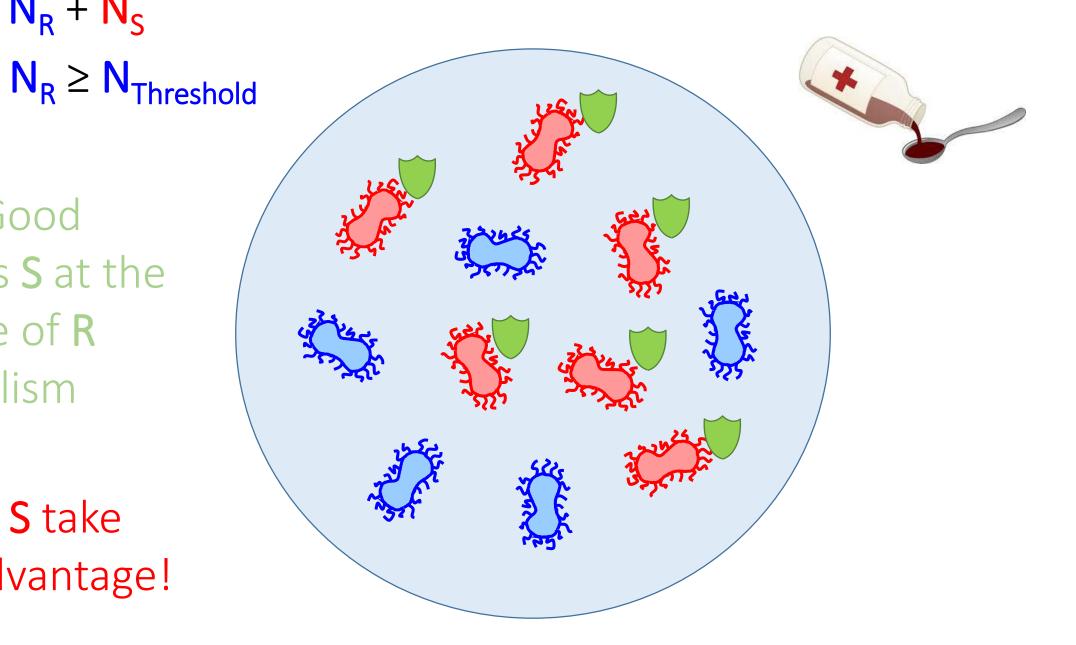
 $N = N_{R} + N_{S}$



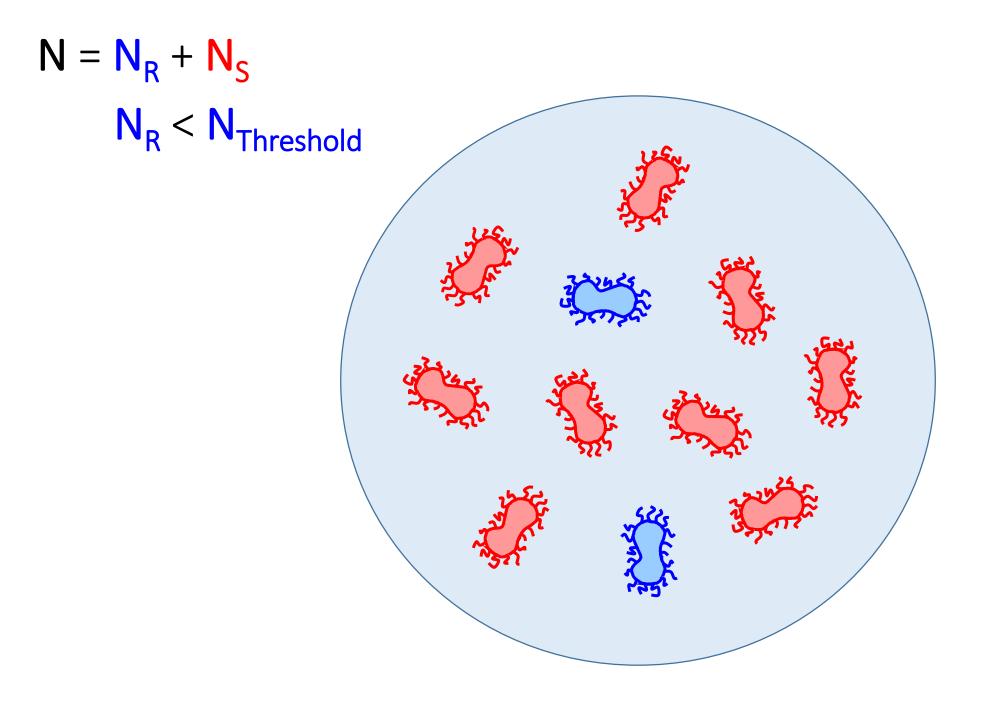
Public Good protects **S** at the expense of R metabolism

 $N = N_{R} + N_{S}$

S take advantage!

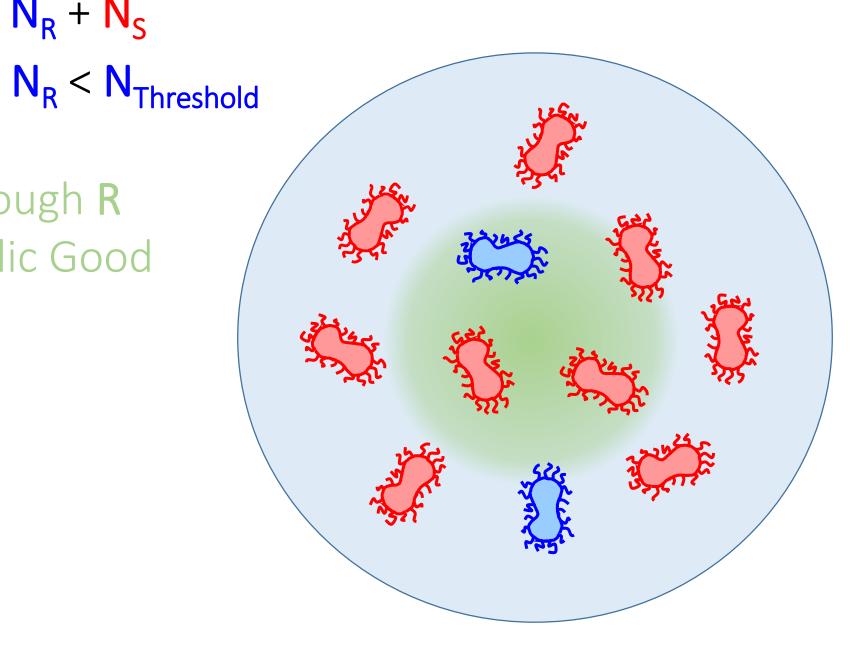


But if $N_R < N_{threshold} \cdots$



Not enough R for Public Good

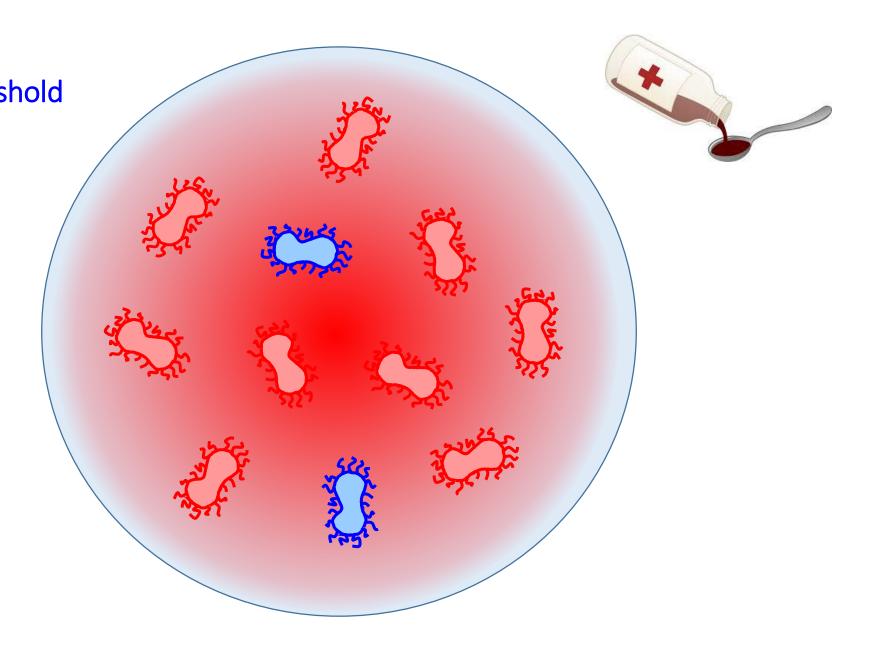
 $N = N_{R} + N_{S}$



 $N = N_{R} + N_{S}$ $N_{R} < N_{Threshold}$ Not enough R

for Public Good

Antimicrobial drug affects **S**

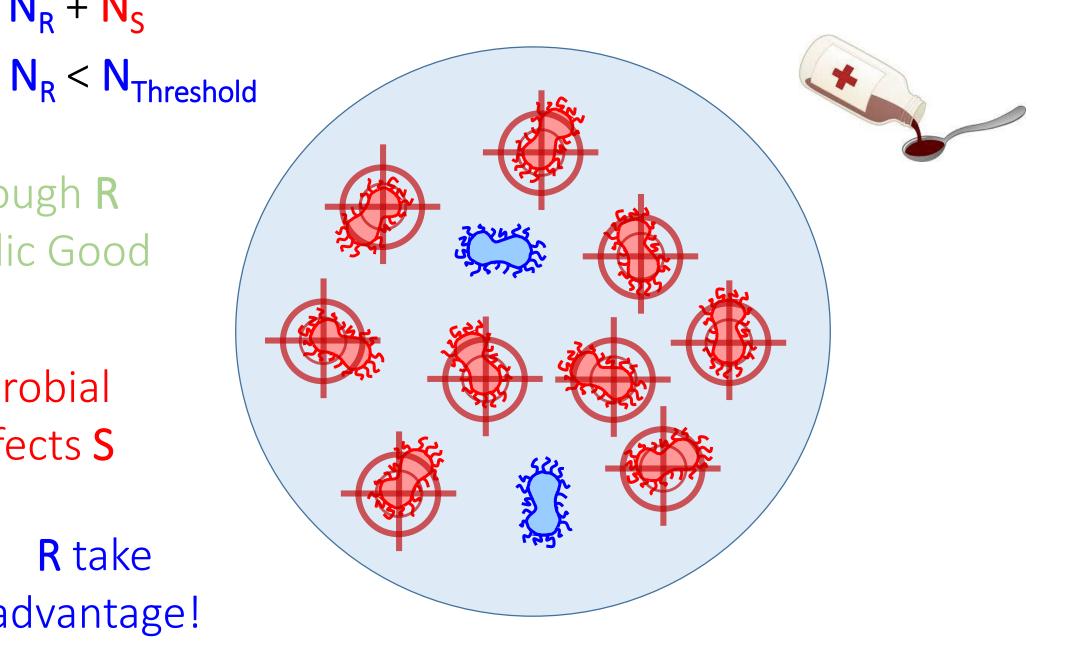


Not enough **R** for Public Good

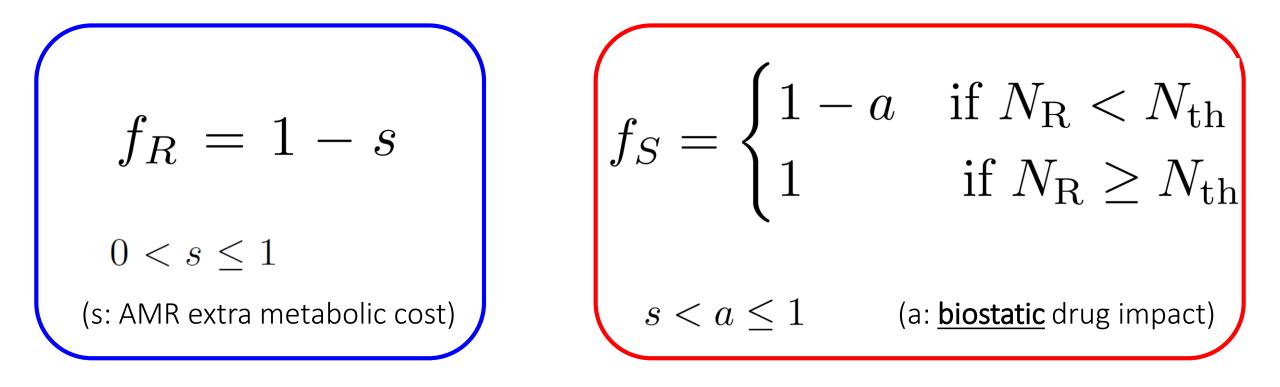
 $N = N_{R} + N_{S}$

Antimicrobial drug affects S

> **R** take advantage!

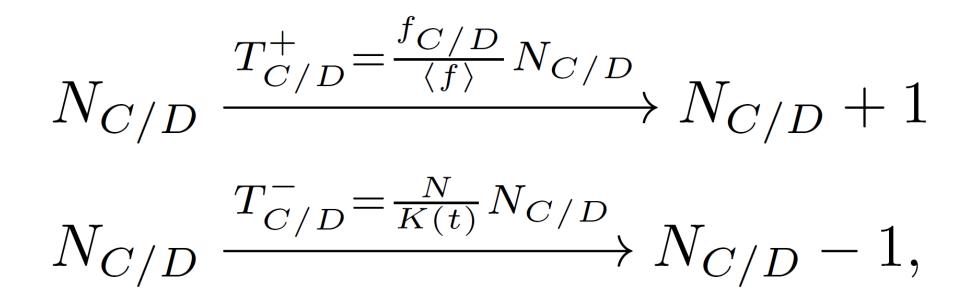


Birth rate per capita $\propto f_{\rm R}$ or $f_{\rm S}$





Transition rates



$$\dot{N} = N\left(1 - \frac{N}{K}\right)$$

• Stable point at N = K

$$x \equiv N_R/N$$

$$\dot{N} = N\left(1 - \frac{N}{K}\right)$$

• Stable point at N = K

 $x \equiv N_R/N$

$$\dot{N} = N\left(1 - \frac{N}{K}\right) \qquad \dot{x} \propto \begin{cases} (a-s) \cdot x(1-x) & 0 \le x < x_{th} \\ \\ -s \cdot x(1-x) & x_{th} \le x \le 1 \end{cases}$$

• Stable point at N = K Equilibrium at x = $x_{th} \equiv N_{th}/N$

(coexistence)

 $x \equiv N_R/N$

Big populations in static environments: Stable coexistence at $N_R = N_{th}$ and $N_S = K - N_{th}$

• Stable point at N = K

• Equilibrium at
$$x = N_{th}/N$$

(coexistence)

Beyond Mean Field:

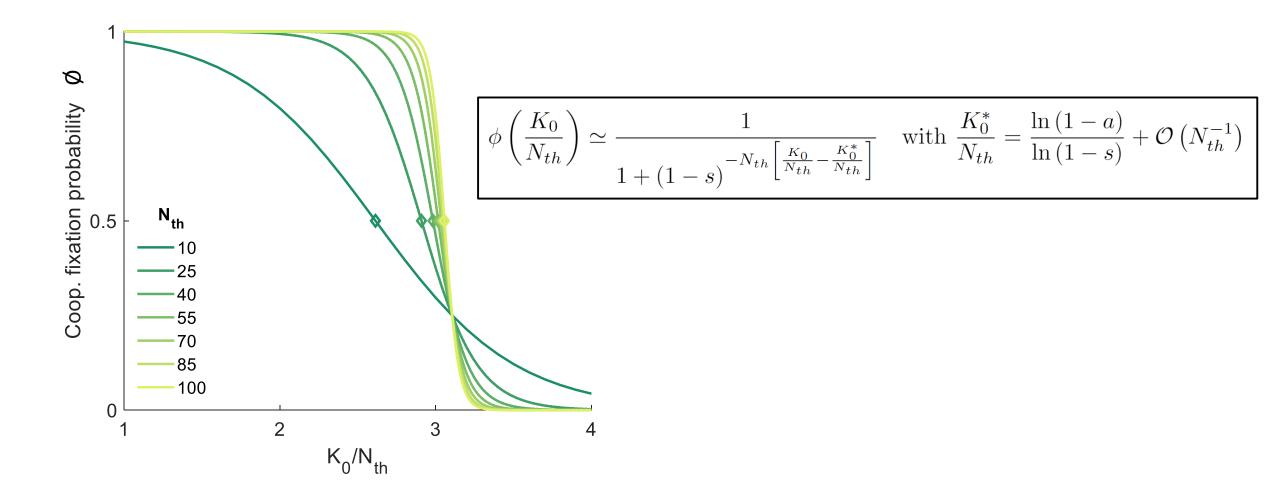
Role of demographic (x) fluctuations

Beyond Mean Field:

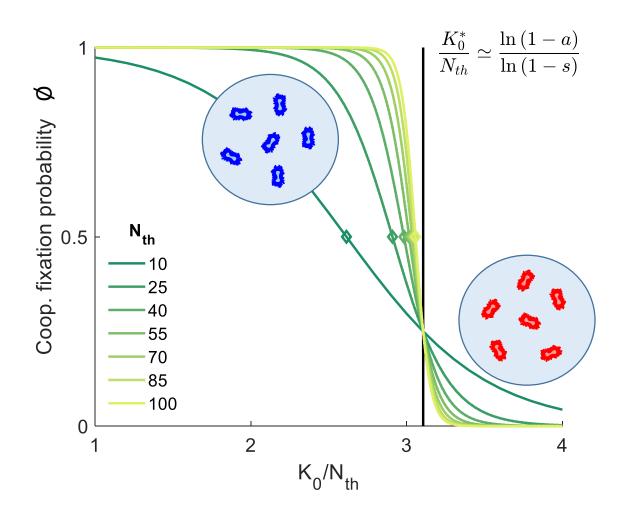
Role of demographic (x) fluctuations

Assume Moran process (fixed $N=K_0$)

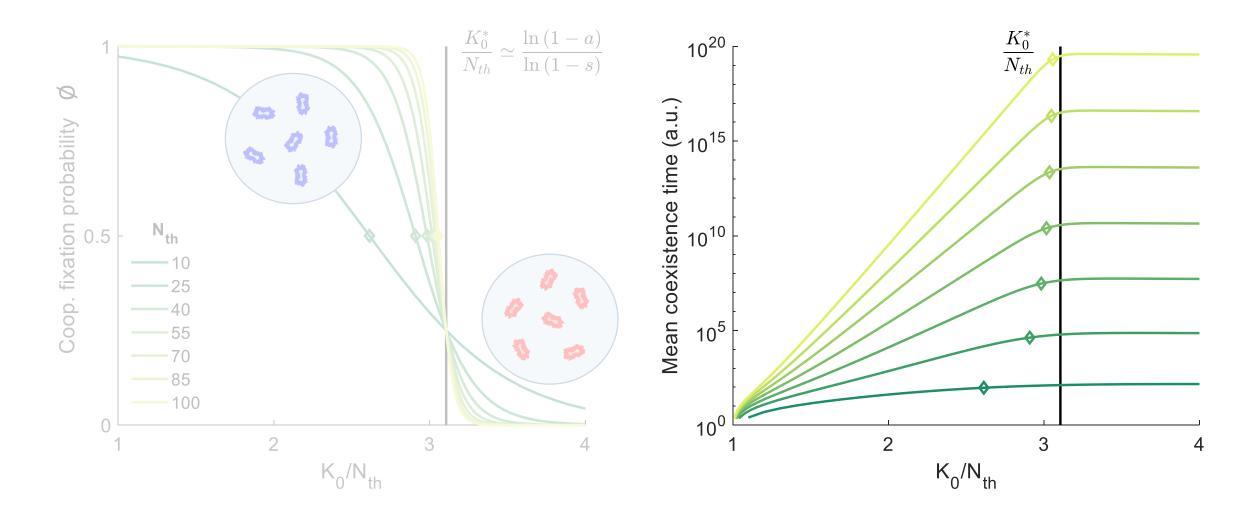
Fixation probability



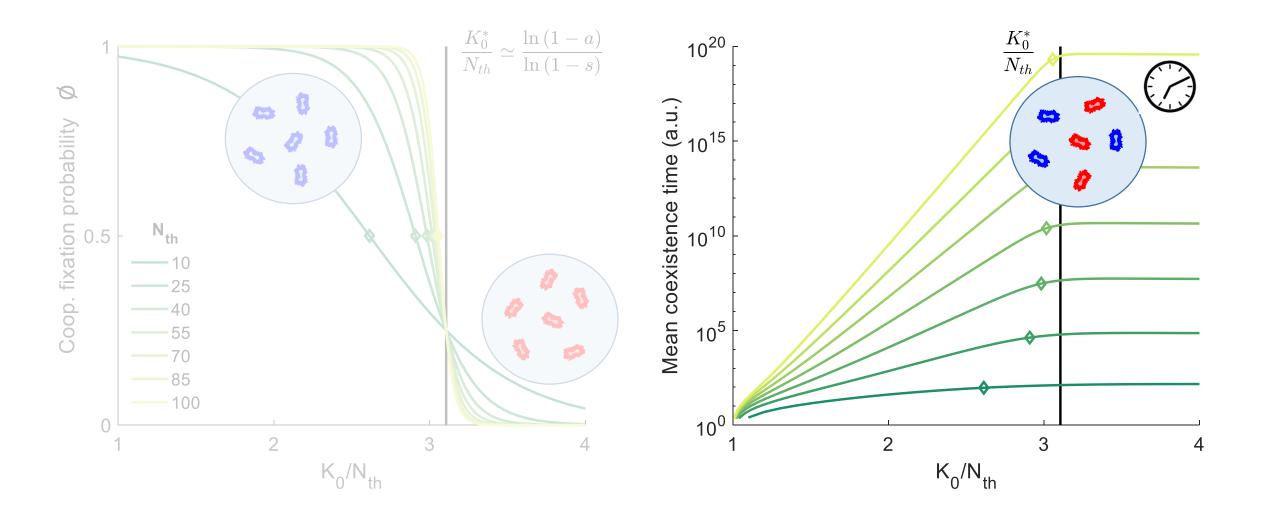
Fixation probability



Coexistence time



Coexistence time



Small populations in static environments:

AMR is doomed to survive!

Does AMR survive in dynamic environments as well?

Eco-Evolutionary dynamics: beyond static environments

Before: demographic fluctuations only

Now: demographic + environmental fluctuations (N changes driven by K(t))

INTERFACE

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Research

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Eco-evolutionary dynamics of cooperative antimicrobial resistance in a population of fluctuating volume and size

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Abstract



Antimicrobial resistance to drugs (AMR), a global threat to human and animal health, is often regarded as resulting from cooperative behaviour. Moreover, microbes generally evolve in volatile environments that, together with demographic fluctuations (birth and death events), drastically alter population size and strain survival. Motivated by the need to better understand the evolution of AMR, we study a population of time-varying size consisting of two competing strains, one drug-resistant and one drug-sensitive, subject to demographic and environmental variability. This is modelled by a binary carrying capacity



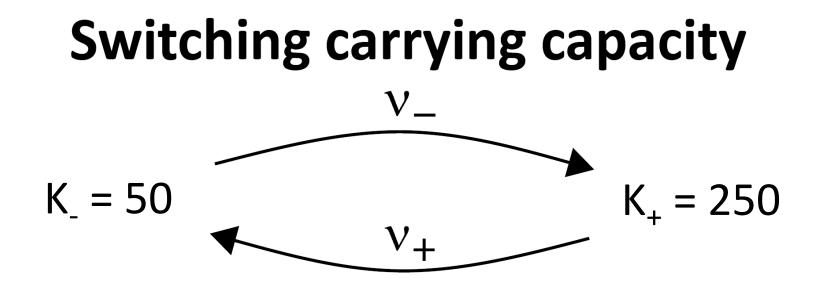
PhD candidate Matthew Asker





Prof. Mauro Pro Mobilia M. al fluctuations driven by K(t))

Prof. Alastair M. Rucklidge



Mean switching rate

$$v \equiv \frac{v_- + v_+}{2}$$

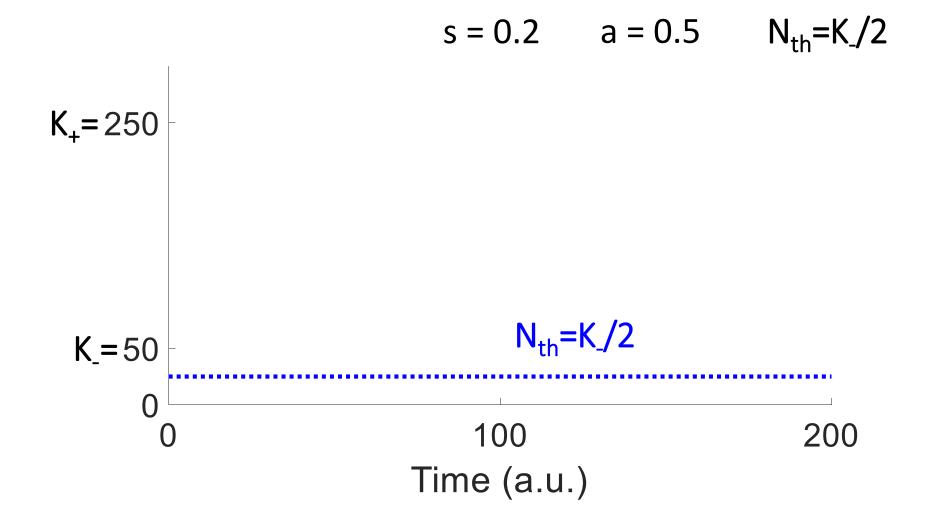
Environmental bias

$$\delta \equiv \frac{\nu_- - \nu_+}{2\nu}$$

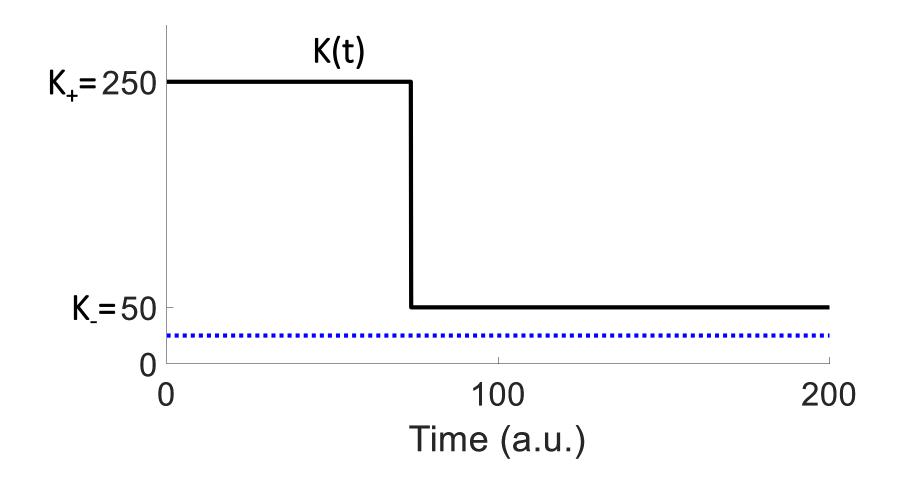
Master equation in dynamic environments

$$\frac{\partial P(N_C, N_D, \xi, t)}{\partial t} = \left(\mathbb{E}_C^- - 1\right) \left[T_C^+ P(N_C, N_D, \xi, t)\right] + \left(\mathbb{E}_D^- - 1\right) \left[T_D^+ P(N_C, N_D, \xi, t)\right] \\ + \left(\mathbb{E}_C^+ - 1\right) \left[T_C^- P(N_C, N_D, \xi, t)\right] + \left(\mathbb{E}_D^+ - 1\right) \left[T_D^- P(N_C, N_D, \xi, t)\right] \\ + \nu_{-\xi} P(N_C, N_D, -\xi, t) - \nu_{\xi} P(N_C, N_D, \xi, t)$$

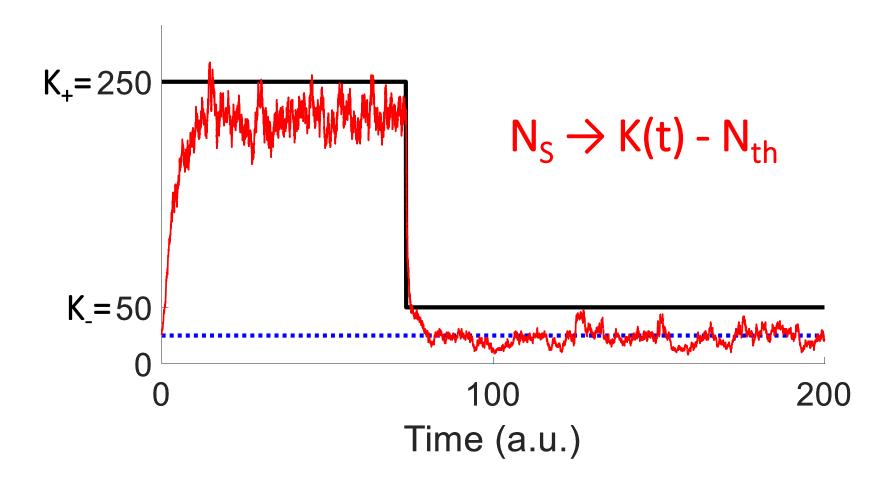
$$\mathbb{E}_{C/D}^{\pm} f(N_{C/D}, N_{C/D}, t) = f(N_{C/D} \pm 1, N_{D/C}, t)$$



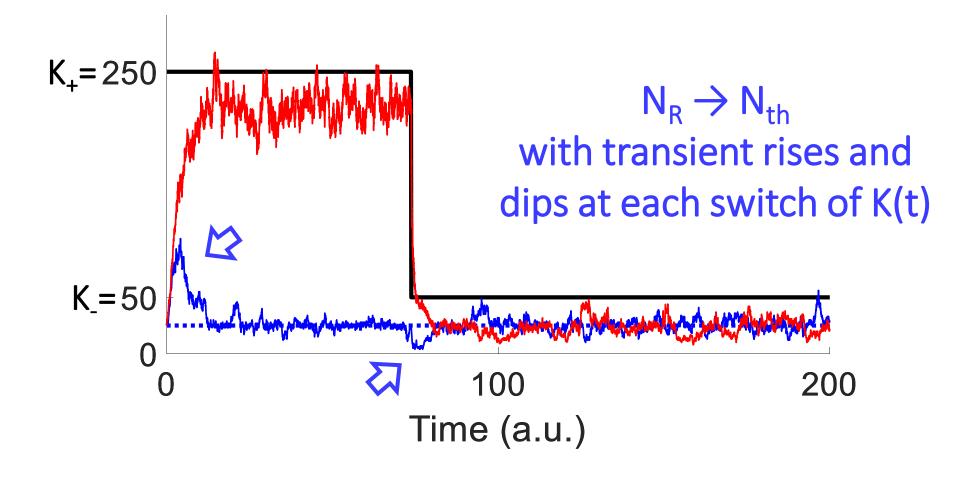
Trajectories in dynamic environments



Trajectories in dynamic environments



Trajectories in dynamic environments



Trajectories in dynamic environments

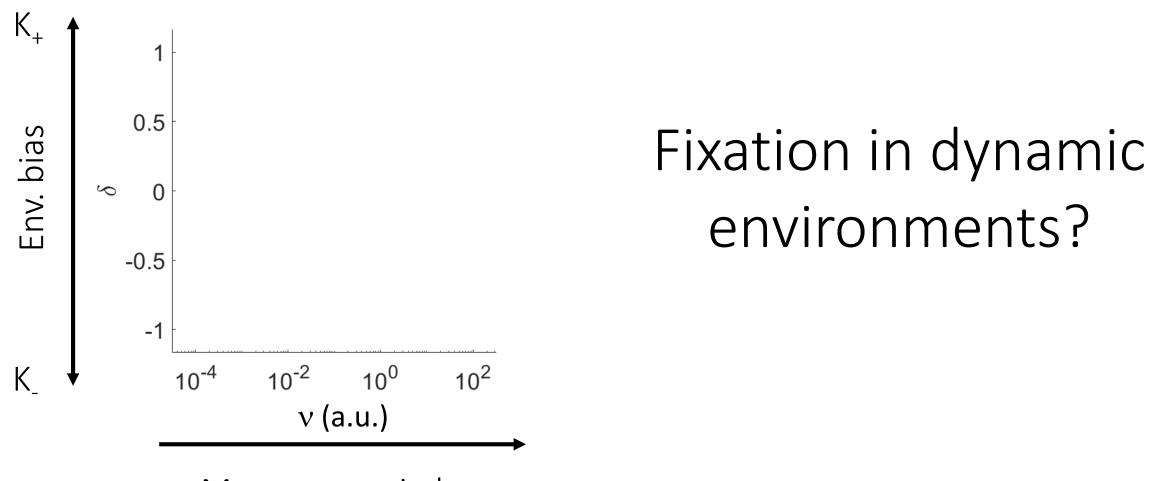
Remember static environments:

Resistant microbes fixate for K_

Sensitive cells fixate for K₊

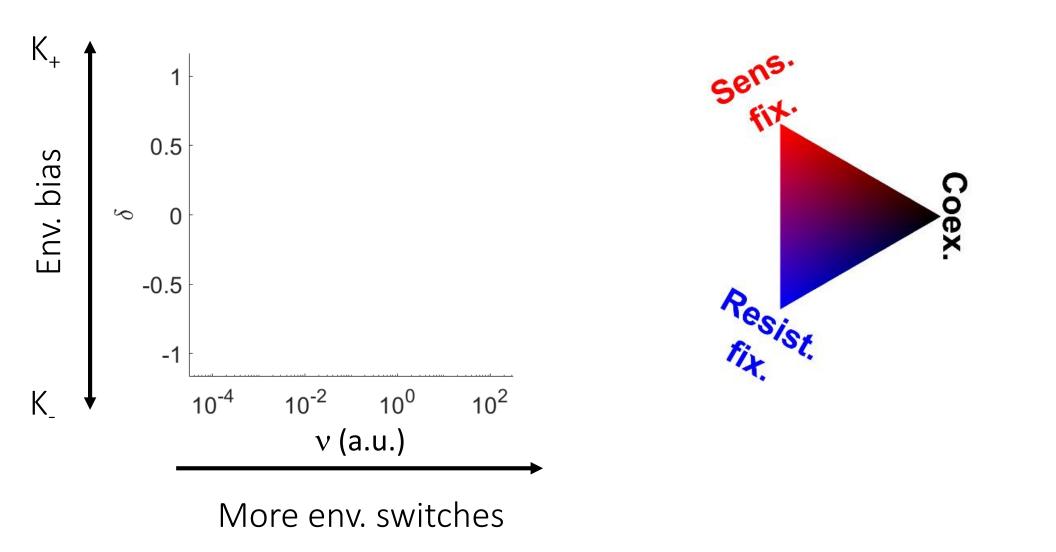
But fixation takes exponentially longer the higher K

s = 0.1 a = 0.25 K₋=120 K₊=1000



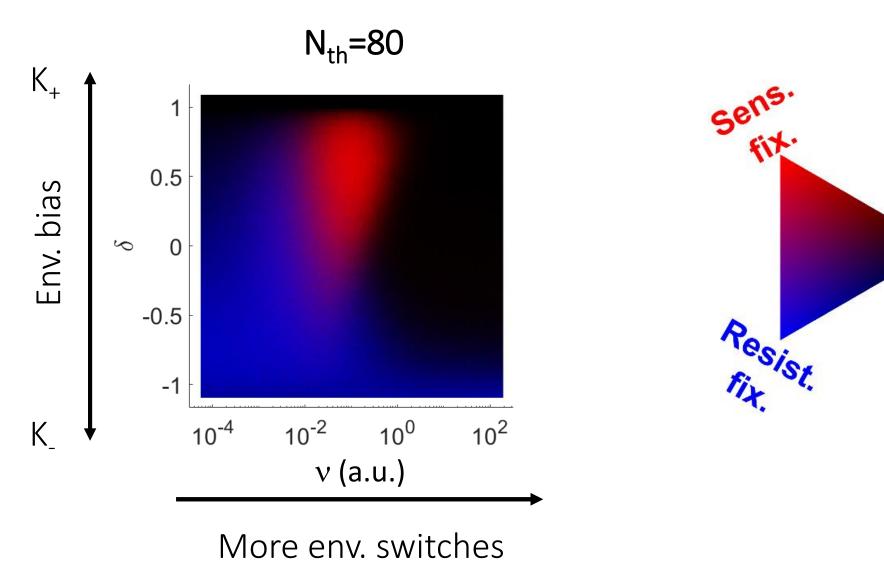
More env. switches

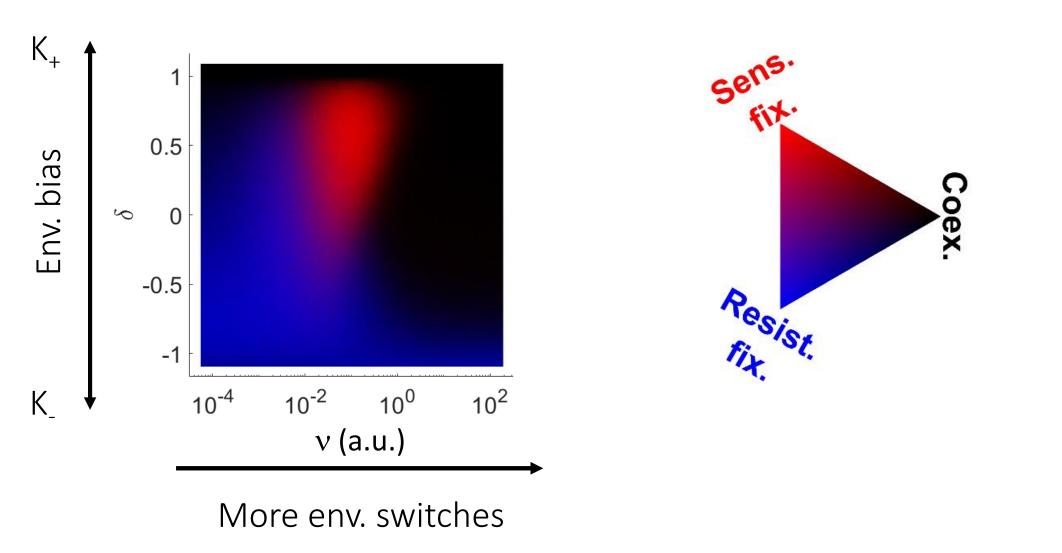


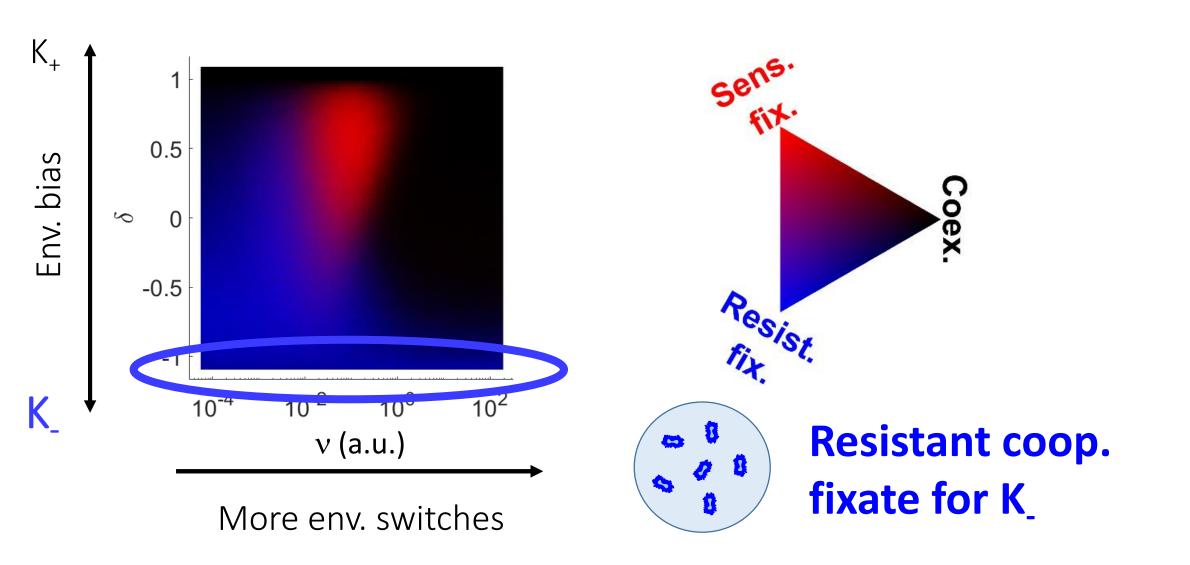


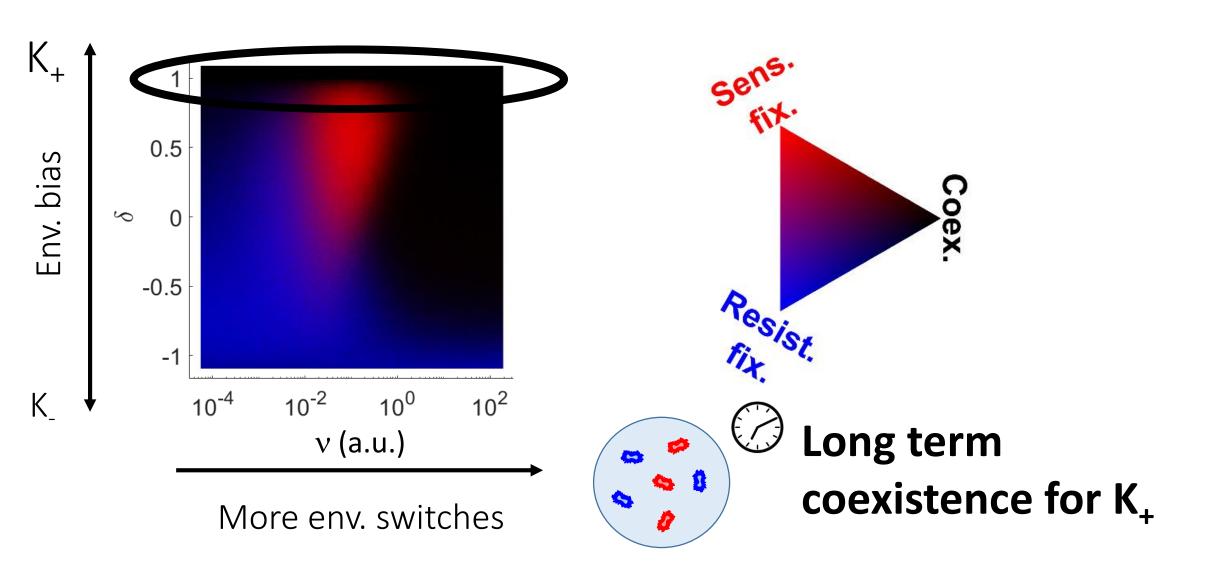
s = 0.1 a = 0.25 K_=120 K_=1000

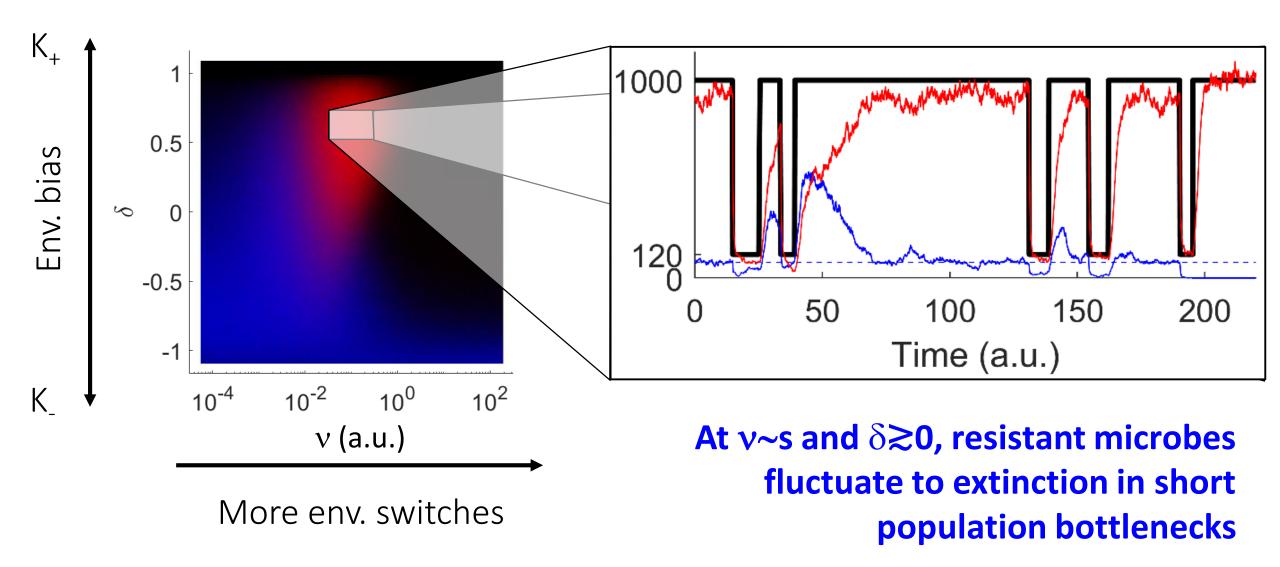
Coex.



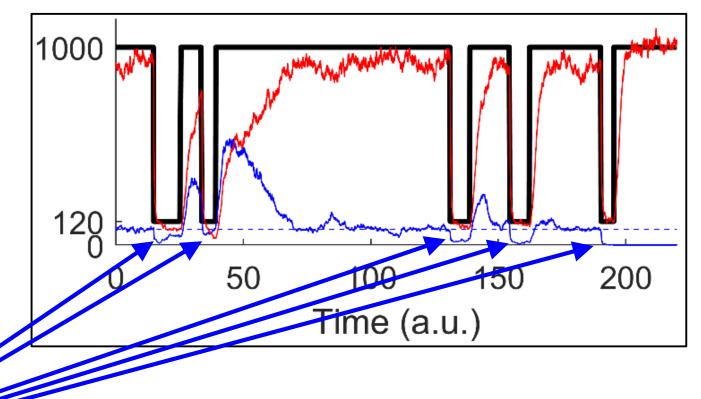


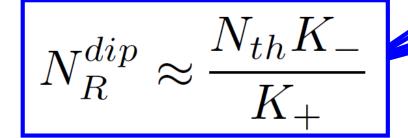






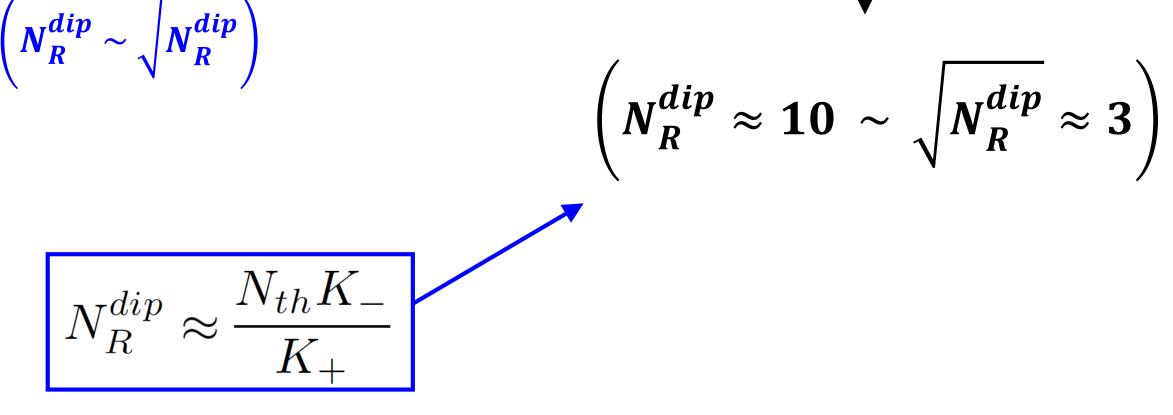
If N_R^{dip} is small enough, then demographic noise is strong enough to drive *R* to extinction $\left(mean\left(N_R^{dip}\right) \sim std\left(N_R^{dip}\right)\right)$





K_=120 K_=1000 N_{th}=80

If N_R^{dip} is small enough, then demographic noise is strong enough to drive *R* to extinction



This fluctuation-driven mechanism works for realistically big microbial populations too!

(e.g., try
$$m N_{th} = 10^{6}$$
, $m K_{-} = 2 \cdot 10^{6}$, and $m K_{+} = 10^{12}$)

$$N_R^{dip} \approx \frac{N_{th}K_-}{K_+}$$

Take-home message(s)

In static environments:

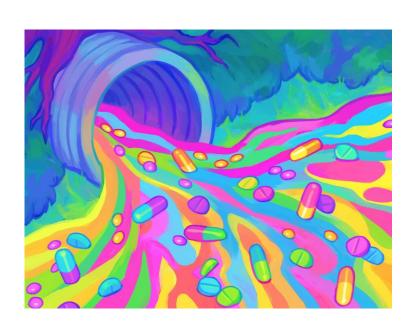
- AMR fixates when public drug-inactivation requires a high proportion of resistant microbes ($N_{th} \approx K$).
- AMR becomes extinct when public drug-inactivation requires a small R fraction ($N_{th} \ll K$), but it usually takes very long.

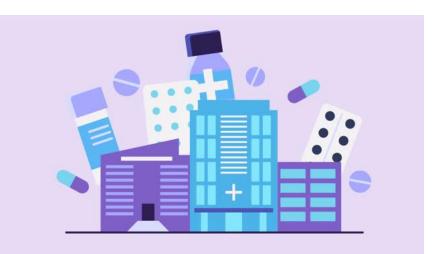
In switching environments:

• Intermediate switching frequencies $(v \sim s)$ enforce and speed up the eradication of AMR through transient dips (if $\frac{N_{th}K_{-}}{K_{+}} \leq O(1)$).

AMR in environments with <u>spatial structure</u> (farms, sewerage, hospitals...)









PhD candidate Kenneth Distefano



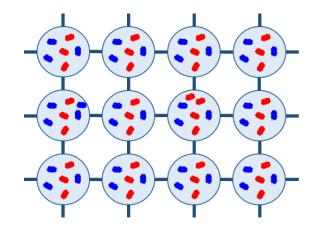
Prof. Uwe C. Täuber



Prof. Mauro Mobilia



AMR fixates/survives in well-mixed, static environments and...

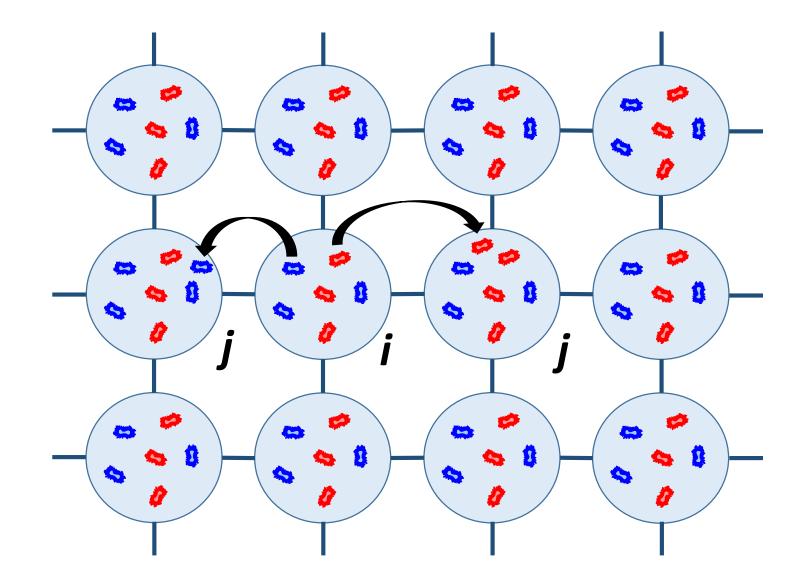


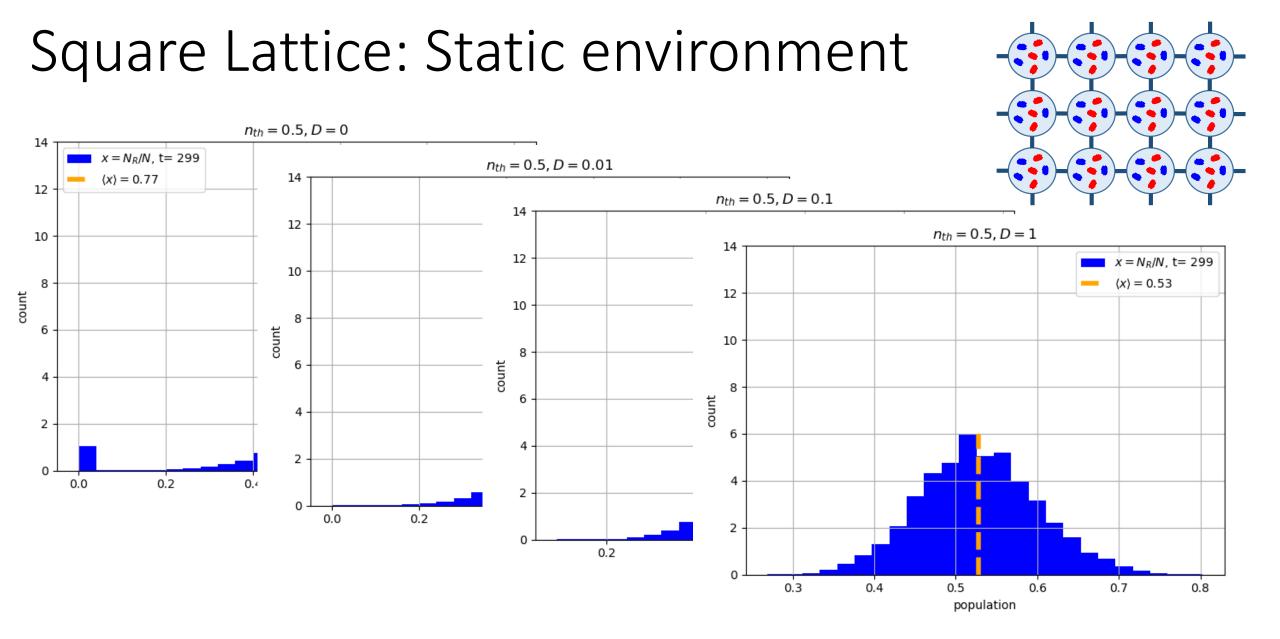
...AMR can be eradicated in dynamic environments but...

...can AMR be eradicated when the environment has spatial structure?

2D periodic square lattice of L x L (gamo)demes

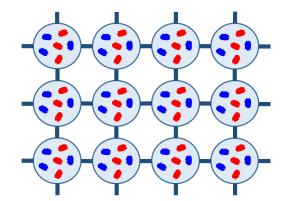
Emigration from deme i to a nearest neighbour random deme (j) at per capita rate: $\mathbf{D} \cdot \mathbf{N}_i / \mathbf{K}(\mathbf{t})$





Spatial migration enforces and shapes strain coexistence

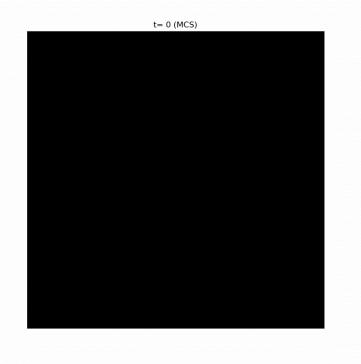
Can we eradicate R?

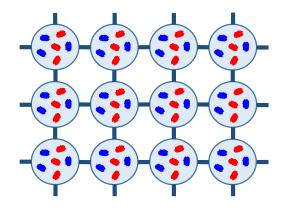


Can we eradicate R?

Frequent Population bottlenecks with high migration rate (D=0.1)

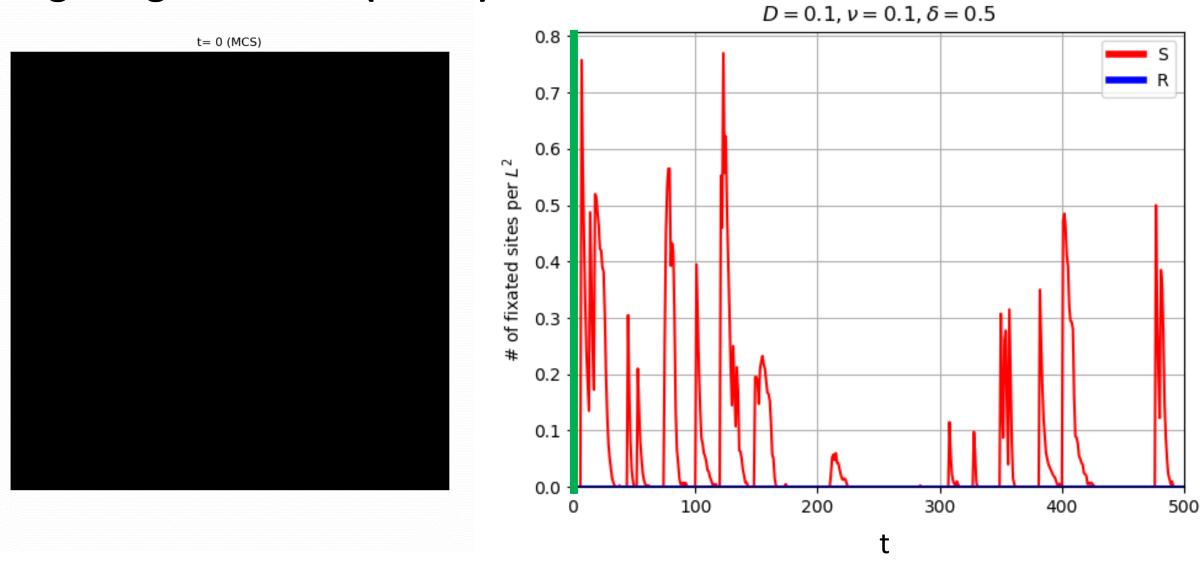
$$L x L = 20 x 20$$





- Site with R and S cells
- Site with no R cells

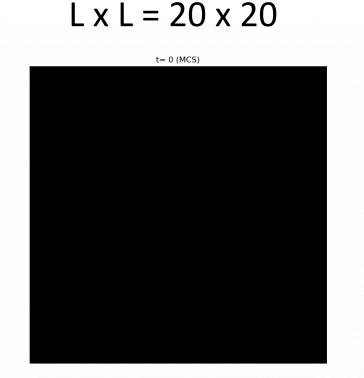
High migration rate (D=0.1)

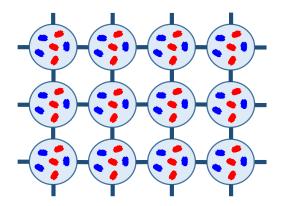


Spatial migration enforces strain coexistence, but...

Can we eradicate R?

Frequent Population bottlenecks with **lower migration rate** (D=0.01)





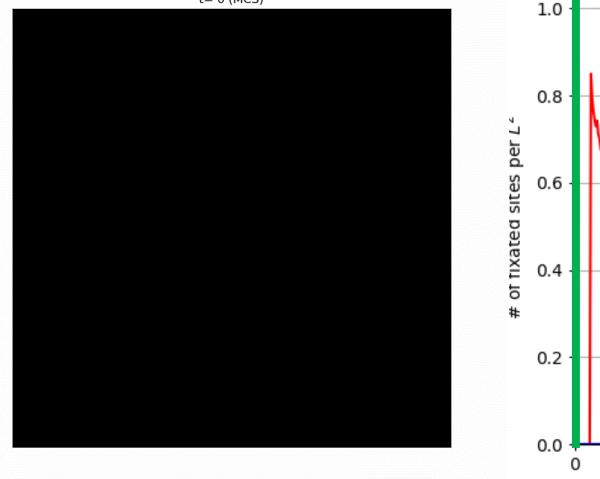
Site with R and S cells

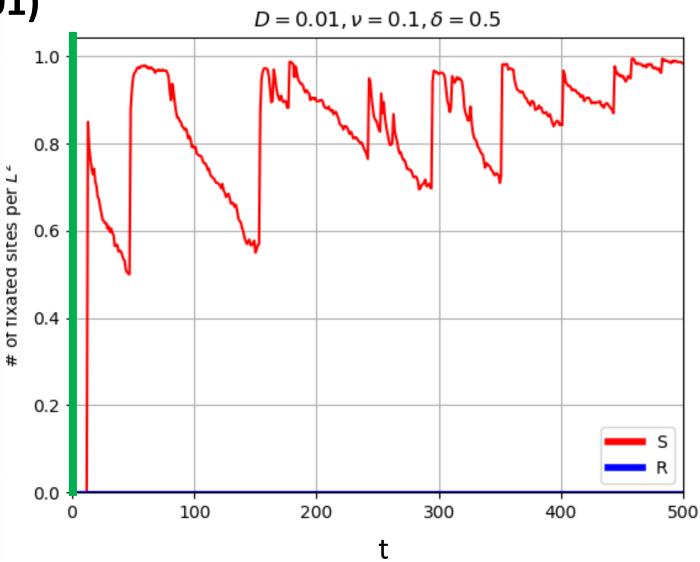
Site with no R cells

...AMR can still be eradicated with strong bottlenecks and/or slow migration!

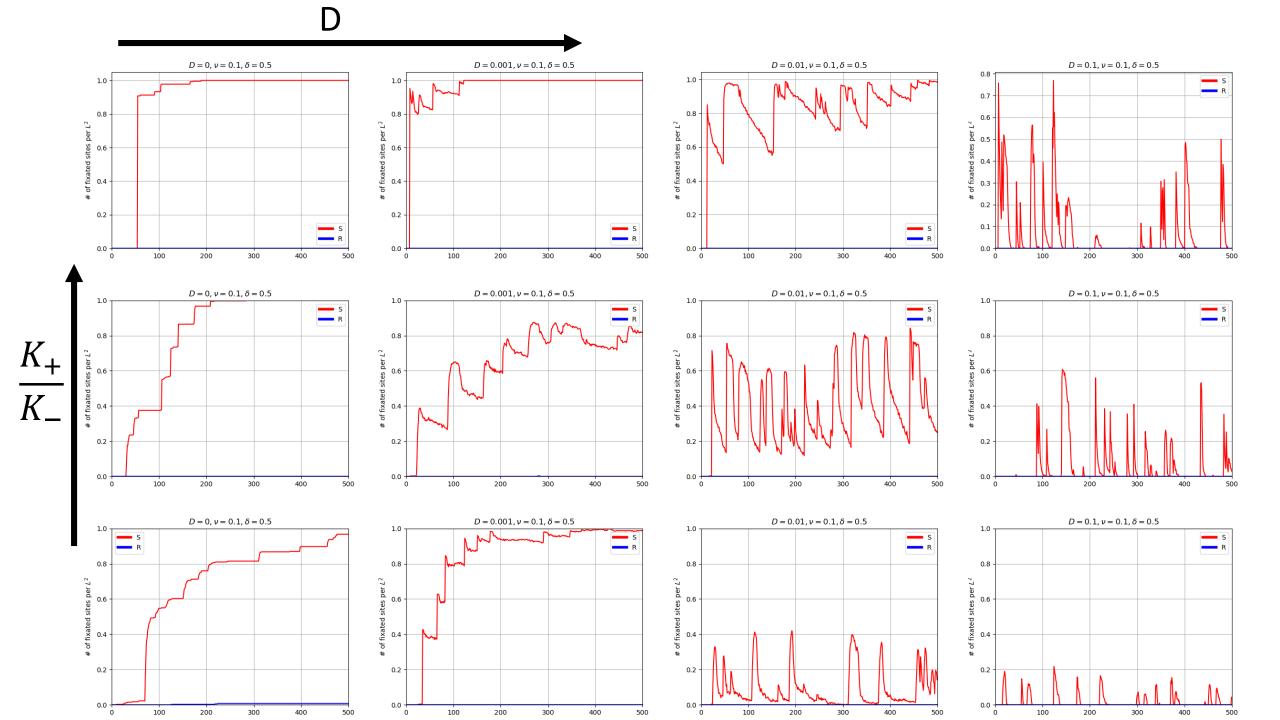
Lower migration rate (D=0.01)

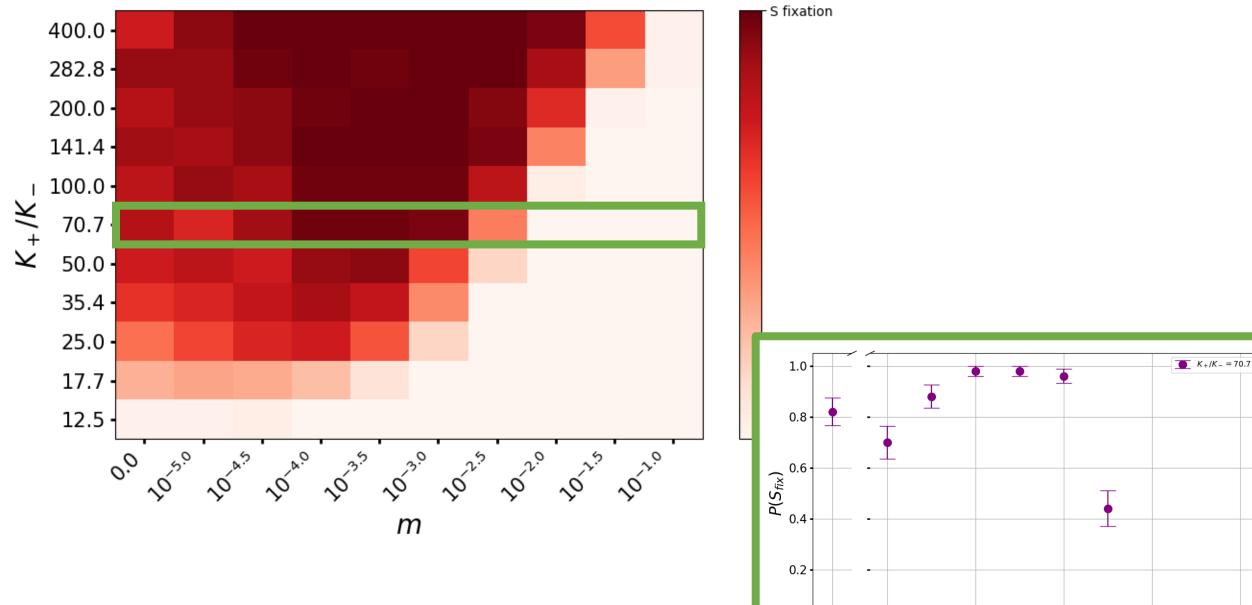
t= 0 (MCS)

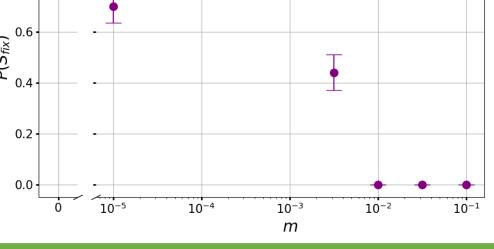




...AMR can still be eradicated!







'Spatial' take-home message(s)

• Faster migration hinders the eradication of AMR.

• But strong population **bottlenecks can still eradicate AMR**.

• And slow-but-non-zero enhances **AMR eradication**.





Engineering and Physical Sciences Research Council



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PhD candidate Said Muñoz Montero



Prof. Uwe C. Täuber



Prof. Michel Pleimling

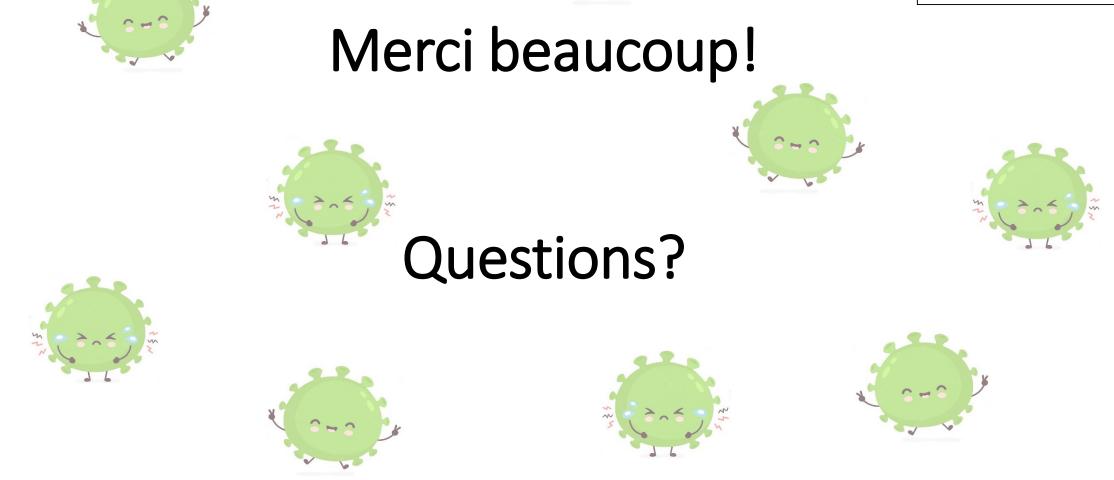


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Coupled Environmental and Demographic Fluctuations Shape the Evolution of Cooperative Antimicrobial Resistance; Hernández-Navarro, L., Asker, M., Rucklidge, A.M., & Mobilia, M.; J. R. Soc. Interface (2023)